

Aquatic Plant-Based Biostimulants: Comparative Evaluation of *Sargassum* sp. and *Eichhornia crassipes* Extracts for Rice Growth Promotion

Husni Asmi¹, Iradhatullah Rahim^{1*}, Syamsiar Zamzam¹, Sukmawati¹, Yuliarti Ramli², Jasman³

¹Faculty of Agriculture, Animal Husbandry, and Fisheries, Universitas Muhammadiyah Parepare, Indonesia

²Faculty of Teacher Training and Education, Universitas Muhammadiyah Parepare, Indonesia

³Faculty of Engineering, Universitas Muhammadiyah Parepare, Indonesia

ORCID/Email:

First AUTHOR : husniasmi11@gmail.com

Second AUTHOR : <https://orcid.org/0000-0002-3913-9611> / iradhat76@gmail.com

Third AUTHOR : <https://orcid.org/0000-0003-2979-7837> / syamsiarzamzam13@gmail.com

Fourth AUTHOR : <https://orcid.org/0000-0001-7452-7110> / sukmakuuh76@gmail.com

Fifth AUTHOR : <https://orcid.org/0009-0008-2303-0663> / yuliartiramli.yr@gmail.com

Sixth AUTHOR : <https://orcid.org/0000-0001-5680-7565> / jasmanyusuf70@gmail.com

Corresponding author email: iradhat76@gmail.com

Article Info

Received: 10/04/2026

Revised: 18/05/2026

Accepted: 20/06/2026

Online: 30/06/2026

Abstract

Aquatic weeds are abundant biomass resources, yet their conversion into compost extracts for rice establishment remains insufficiently evaluated across the transition from seed germination to nursery growth. This study examined compost extracts from *Sargassum* sp. and water hyacinth (*Eichhornia crassipes*) as stage-specific nutrient-biostimulant inputs for *Oryza sativa* L. A randomized block design was used for germination, combining four extract types (biohumic, *Sargassum* sp., water hyacinth, and their mixture) with four concentrations (25, 50, 75, and 100 mL L⁻¹). Extract chemistry and treatment means were interpreted conservatively because ANOVA did not detect significant treatment effects. The results showed *Sargassum* sp. extract had higher organic C, C/N ratio, K-equivalent concentration, and pH, whereas water hyacinth had higher N. Biohumic at 25 mL L⁻¹ showed the highest germination means for plumule length, radicle length, leaf number, and fresh mass. The results support a cautious, hypothesis-generating model in which extract performance depends on rice developmental stage. Low-dose humic input appears more suitable for germination, whereas higher-dose aquatic weed extracts may be more useful during nursery growth.

Keywords: *Oryza sativa*; *Sargassum* sp.; water hyacinth; compost extract; liquid organic biostimulant



© 2026 by the author(s)

This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

INTRODUCTION

Rice (*Oryza sativa* L.) establishment is a critical phase in crop production because seed germination, root initiation, early leaf expansion and nursery-stage seedling vigor determine stand uniformity and the capacity of plants to respond to later fertilization. Nitrogen is central to these processes because it contributes to chlorophyll formation, protein synthesis, photosynthetic activity and

plant architecture, including root development, leaf expansion, tillering and biomass accumulation (Luo et al., 2020; Qi et al., 2025; Wang et al., 2024). However, fertilizer strategies for rice increasingly need to combine productivity with reduced dependence on mineral fertilizer and improved recycling of organic resources (Barus et al., 2024; Haque et al., 2021; Kaur et al., 2023; Marti-Jerez et al., 2023).

Organic and bio-organic inputs can improve rice growth, nutrient uptake and soil biological activity, but their effects are not universal. Recent studies on liquid organic fertilizers, humic inputs, microbial biostimulants and organic amendments indicate that crop response depends on feedstock chemistry, dose, plant developmental stage and environmental context (Cabrera et al., 2025; Chen et al., 2025; Dwi Nurcahyanti et al., 2024; Iriany et al., 2023; Riddech et al., 2025). For early rice establishment, this conditionality is especially important because germinating seeds are more sensitive than established seedlings to pH, soluble salts, oxygen limitation and potentially phytotoxic dissolved organic compounds.

Aquatic weeds provide a biologically and environmentally relevant feedstock for circular agricultural input development. *Sargassum* sp. is a brown marine macroalga that contains mineral and carbon-rich biochemical fractions, while water hyacinth (*Eichhornia crassipes*) is a rapidly proliferating freshwater macrophyte that can accumulate nutrients and generate large unmanaged biomass (Asmara et al., 2020; Begum et al., 2022; Choi et al., 2025; Liu et al., 2025; Zega et al., 2025). Converting these biomass streams into compost extracts may reduce disposal pressure and provide locally available inputs for rice nurseries. Yet the value of such extracts cannot be inferred from nutrient content alone, because seaweed-derived and aquatic-plant-derived products may also contain bioactive or inhibitory organic fractions (Banakar et al., 2022; Banakar et al., 2023; Fitriyah et al., 2022; Senthilkumar et al., 2024; Utomo & Guntoro, 2023; Xie et al., 2021).

Humic inputs provide a useful benchmark for this question. Humic acid and humic-like substances have been associated with improved rice germination, metabolic adjustment, rhizosphere nutrient distribution and field performance, although their effects also depend on stress context and formulation (Adhikari et al., 2023; Canellas et al., 2022; Ibraheem et al., 2023; Indriani et al., 2022; Izquierdo et al., 2025; Ji et al., 2024; Minhas et al., 2024). Comparing aquatic weed extracts with a humic benchmark therefore makes it possible to ask whether these local biomass-derived extracts act similarly across establishment stages or whether their usefulness is stage-dependent.

The research gap addressed here is not simply whether aquatic weed biomass contains nutrients, but whether compost extracts from contrasting aquatic weeds show different response profiles during seed germination and seedling growth. The study was designed to evaluate nutrient composition, germination response and nursery-stage growth response of rice exposed to biohumic, *Sargassum* sp., water hyacinth and combined extracts. In response to reviewer concerns, the present revised manuscript frames the results as a preliminary stage-specific response model rather than as definitive evidence of treatment superiority, because the available ANOVA results did not indicate statistically significant treatment effects.

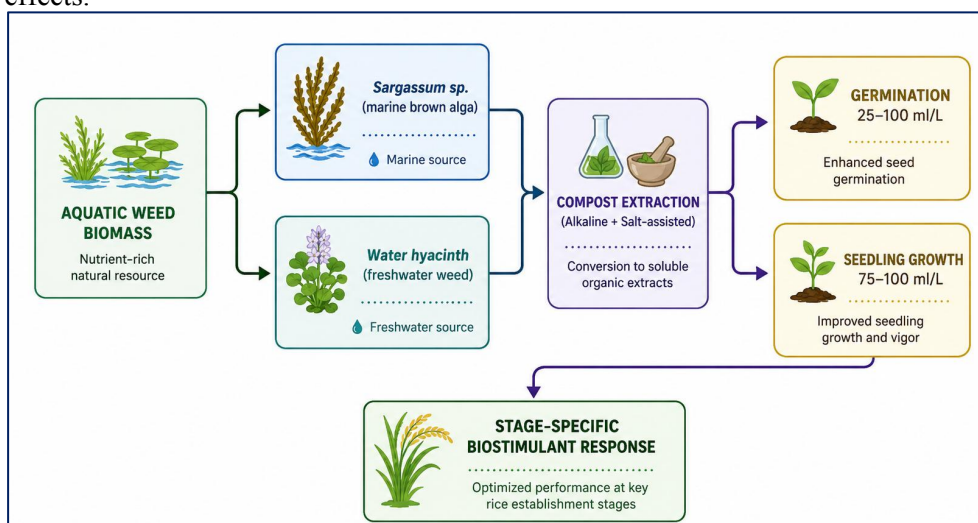


Figure 1. Experimental logic linking aquatic weed biomass, compost extraction, concentration gradient and stage-specific rice establishment responses

RESEARCH METHOD

Study site and biological material

The experiment was conducted at the Integrated Laboratory of the Faculty of Agriculture, Animal Husbandry and Fisheries, Universitas Muhammadiyah Parepare, Indonesia, from July 2025 to April 2026. The test crop was rice (*Oryza sativa* L.) cultivar Inpari. The aquatic materials were composted *Sargassum* sp. and composted water hyacinth (*Eichhornia crassipes*), representing contrasting marine and freshwater weed biomass sources. Biohumic was included as a reference organic biostimulant to compare aquatic weed extracts with a humic-type benchmark.

Compost extract preparation

Composted aquatic weed material was sun-dried, ground and sieved to obtain a fine powder. For each extract, 100 g of powdered compost was mixed with 1000 mL distilled water. NaOH was dissolved into the aqueous phase before the compost powder was added, and the suspension was homogenized using a magnetic stirrer for 2 h. The suspension was allowed to stand for 24 h, NaCl solution was added, and the mixture was left overnight to improve separation. The liquid fraction was separated from sediment, centrifuged at 4000 rpm for 15 min, and filtered through Whatman 42 paper to obtain a clarified extract. The procedure was intended to recover soluble nutrient and humic-like fractions while removing coarse particles that could interfere with seed contact and oxygen diffusion (Du et al., 2023; Kurniawan et al., 2024; Ros et al., 2020).

Experimental design

The germination experiment used a factorial randomized block design with two factors. The first factor was extract type: biohumic (N1), *Sargassum* sp. extract (N2), water hyacinth extract (N3), and combined *Sargassum* sp. + water hyacinth extract (N4). The second factor was extract concentration: 25, 50, 75 and 100 mL L⁻¹ (K1-K4). The 4 x 4 factorial structure produced 16 treatment combinations arranged in three blocks, giving 48 experimental units.

Measurements

During germination, treated seeds were placed on merang paper and evaluated at week 5. Plumule length was measured from the shoot base to the highest leaf tip, radicle length from the root base to the longest root tip, and root and leaf numbers were counted directly. Fresh mass was recorded immediately after harvest, and dry mass was recorded after oven drying. During seedling growth, plant height, leaf number, tiller number, root length, root number, fresh mass and dry mass were evaluated at week 8. These traits describe shoot emergence, root initiation, vegetative vigor, water accumulation and dry matter formation.

Chemical analysis and terminology

Extract chemistry was assessed for organic C, N, C/N ratio, P, K and pH. The source dataset reported phosphate and potassium oxide equivalent values; these were converted to elemental P and K equivalents for journal terminology using standard conversion factors ($P = P_2O_5 \times 0.4364$; $K = K_2O \times 0.8301$). This terminology follows the Plant and Soil preference for elemental nutrient reporting. Because additional indicators such as electrical conductivity, nitrate, ammonium, dissolved organic C, total dissolved solids and phenolics were not measured, mechanistic interpretation is explicitly treated as provisional.

Statistical interpretation

The source analysis used ANOVA followed by a post hoc test when appropriate, and reported that treatment effects were not statistically significant for the measured variables. Complete raw replicate-level data, F-values, degrees of freedom, p-values and standard errors were not available in the manuscript file used for revision. Therefore, the revised Results section avoids claims of statistical superiority and presents treatment means as descriptive patterns only. Figures based on normalized values are intended to support visual synthesis and do not imply significant differences. This cautious reporting directly addresses the reviewer concern that mean-based contrasts could otherwise overstate the strength of the evidence.

RESULTS AND DISCUSSION

Extract chemistry

The two aquatic weed extracts differed in chemical profile (Table 1). *Sargassum* sp. extract had higher organic C, C/N ratio, K-equivalent concentration and pH. Water hyacinth extract had higher N, whereas the converted P-equivalent concentration was similar for both extracts. These differences indicate that *Sargassum* sp. contributed a more carbon-rich and K-associated extract, while water hyacinth provided a relatively N-rich and lower-C/N extract. Both extracts were alkaline, which may have influenced seed-solution interactions and nutrient availability.

Table 1. Chemical composition of *Sargassum* sp. and water hyacinth compost extracts used in the rice establishment experiment.

Extract	Organic C (%)	N (%)	C/N ratio	P equivalent (%)	K equivalent (%)	pH
<i>Sargassum</i> sp.	0.58	0.07	8.15	0.0105	0.0232	8.5
Water hyacinth	0.27	0.09	3.07	0.0105	0.0158	8.4

Note: P and K are reported as elemental equivalents converted from the source oxide-equivalent values to align with Plant and Soil terminology.

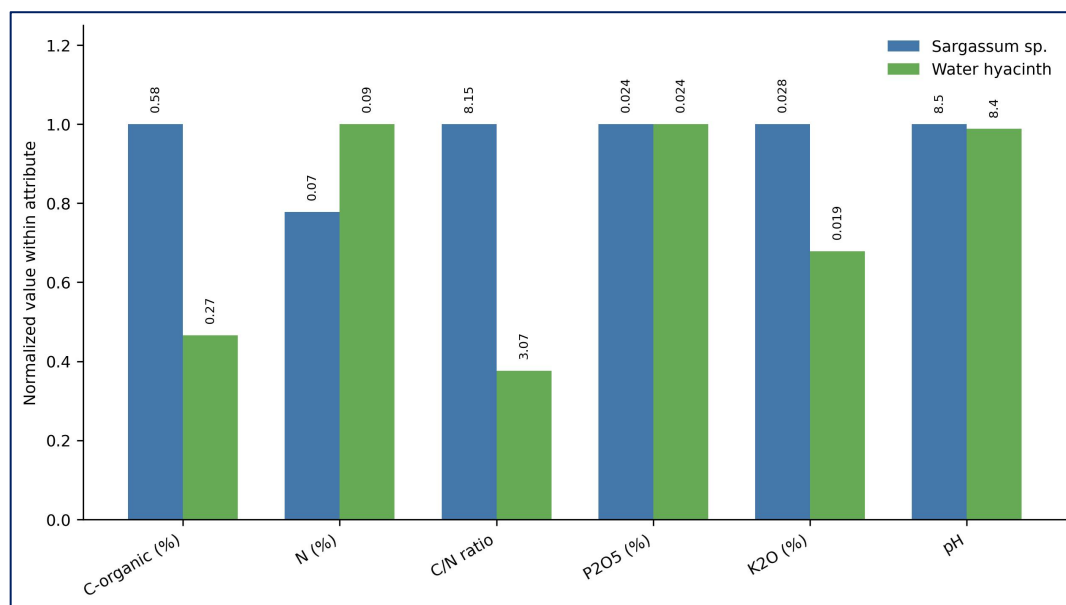


Figure 2. Relative nutrient profile of *Sargassum* sp. and water hyacinth extracts. Values are normalized within each attribute; raw values in the figure correspond to the source extract-chemistry dataset

Germination response

ANOVA did not detect significant treatment effects for germination traits. Nevertheless, the treatment means showed a consistent descriptive pattern. Biohumic at 25 mL L⁻¹ (N1K1) had the highest mean plumule length, radicle length, leaf number and fresh mass, while biohumic at 25 and 50 mL L⁻¹ showed the highest mean root number. Dry mass reached the same maximum mean value in several treatments, including low-dose biohumic, low-dose *Sargassum* sp. and combined extract treatments at 75 and 100 mL L⁻¹. Water hyacinth at 75 mL L⁻¹ (N3K3) had zero mean values for all germination traits. This pattern is reported descriptively and should not be interpreted as statistically proven toxicity.

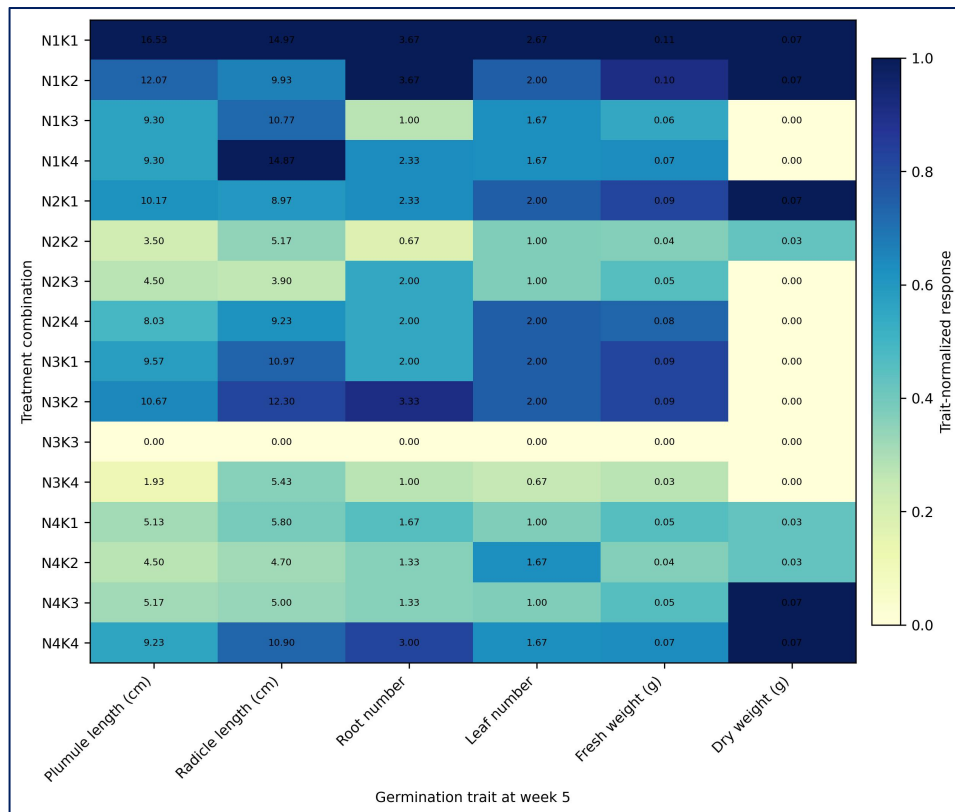


Figure 3. Mean germination responses of rice at week 5 under four extract types and four concentrations. Cell values are treatment means; color intensity indicates trait-normalized descriptive response and does not imply statistical significance.

Descriptive synthesis

The descriptive synthesis suggests a stage-dependent response pattern. During germination, low-dose biohumic showed the most favorable mean profile. During nursery growth, higher-dose aquatic weed extracts showed the most favorable mean profile, with Sargassum sp. associated mainly with height and root number and water hyacinth associated mainly with leaf production, tillering and biomass. Because statistical significance was not detected, this pattern is best interpreted as a preliminary response model that should guide future dose-response and mechanistic experiments rather than as an agronomic recommendation.

Table 2. Highest and lowest treatment means for rice germination responses

Stage	Variable	Highest mean treatment	Highest value	Lowest mean treatment	Lowest value	Inference
Germination	Plumule length (cm)	N1K1	16.53	N3K3	0.00	ANOVA not significant
Germination	Radicle length (cm)	N1K1	14.97	N3K3	0.00	ANOVA not significant
Germination	Root number	N1K1/N1K2	3.67	N3K3	0.00	ANOVA not significant
Germination	Leaf number	N1K1	2.67	N3K3	0.00	ANOVA not significant
Germination	Fresh mass (g)	N1K1	0.11	N3K3	0.00	ANOVA not significant
Germination	Dry mass (g)	Multiple treatments	0.07	Multiple treatments	0.00	ANOVA not significant

Note: N1 = biohumic; N2 = Sargassum sp.; N3 = water hyacinth; N4 = Sargassum sp. + water hyacinth. K1-K4 = 25, 50, 75 and 100 mL L⁻¹. Values are descriptive treatment means.

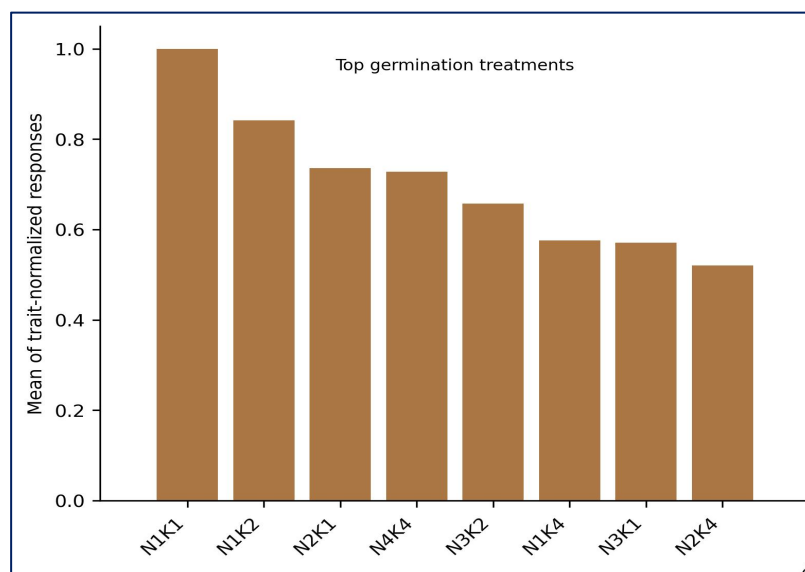


Figure 5. Descriptive integrated vigor indices for germination and seedling growth. The indices are unweighted means of trait-normalized responses and are provided only as exploratory synthesis.

Discussion

Stage-specificity rather than treatment superiority

The strongest contribution of this study is the separation of germination from seedling growth. Rice seeds and established seedlings differ in physiology: germination depends on water imbibition, reserve mobilization, oxygen availability and early hormonal regulation, whereas seedling growth increasingly depends on photosynthesis, nutrient uptake, root branching and tiller initiation (Dai et al., 2025; Ishaq et al., 2025; Linda et al., 2024; Monajjem et al., 2023; Vaishnavi et al., 2024). The observed mean reversal between low-dose biohumic at germination and higher-dose aquatic weed extracts at seedling stage is therefore biologically plausible, but it remains descriptive because the treatment effects were not statistically significant.

Low-dose humic input during germination

Biohumic at 25 mL L⁻¹ showed the highest mean performance for several germination traits. This agrees with the broader literature indicating that humic substances can influence early root development, nutrient availability and rice metabolic adjustment (Canellas et al., 2022; Ibraheem et al., 2023; Indriani et al., 2022; Ji et al., 2024). The response was not linear with concentration, supporting the view that seed-stage biostimulant application requires careful dilution. Higher concentrations may alter water potential, increase dissolved solids or expose seeds to organic fractions that are not favorable during imbibition. Because electrical conductivity and osmotic potential were not measured, this explanation should be treated as a proposed mechanism rather than a demonstrated cause.

Water hyacinth caution at the seed stage

Water hyacinth at 75 mL L⁻¹ had the weakest germination mean profile despite its higher N content. This contrast shows why nutrient content alone is insufficient to predict early seed response. Water hyacinth biomass can improve soil and plant properties after composting, but plant extracts may also contain bioactive fractions with inhibitory potential if decomposition or dilution is unsuitable (Begum et al., 2022; Guidoni et al., 2021; Rosawanti, 2019; Utomo & Guntoro, 2023; Zega et al., 2025). Future work should include a standard germination index, electrical conductivity, total dissolved solids, pH, total phenolics and phytotoxicity screening before recommending direct seed exposure.

Implications for circular rice nursery management

Aquatic weed biomass can be considered a candidate bioresource for circular rice nursery inputs, but formulation science is essential. The present results do not support a single universal concentration. Instead, they support a cautious two-step hypothesis: a low-dose humic or highly diluted extract treatment for germination, followed by stronger Sargassum sp. or water hyacinth extract during nursery growth. This approach aligns with integrated nutrient-management and waste-valorization

studies showing that organic inputs can improve rice performance when feedstock, dose and crop demand are synchronized (Bagheri Novair et al., 2024; Barus et al., 2024; Cabrera et al., 2025; Consorti et al., 2025; Dwi Nurcahyanti et al., 2024; Haque et al., 2021; Kaur et al., 2023; Kanga et al., 2026; Kumar et al., 2024; Marti-Jerez et al., 2023; Prasedya et al., 2023; Riddech et al., 2025; Rozen et al., 2024; Selvam & Kumar, 2022; Thaimei et al., 2025; Umamaheswari & Shanthakumar, 2021; Zhang et al., 2025).

CONCLUSION

This study evaluated compost extracts from *Sargassum* sp. and water hyacinth as potential inputs for rice establishment. *Sargassum* sp. extract had higher organic C, C/N ratio, K-equivalent concentration and pH, while water hyacinth extract had higher N. ANOVA did not detect significant treatment effects; therefore, the findings should be interpreted as descriptive and hypothesis-generating. The highest germination means were associated mainly with biohumic at 25 mL L⁻¹, whereas the highest seedling-growth means shifted toward 100 mL L⁻¹ aquatic weed extracts. *Sargassum* sp. at 100 mL L⁻¹ showed the highest mean plant height and root number, while water hyacinth at 100 mL L⁻¹ showed the highest mean leaf number, tiller number, root length and biomass.

The revised interpretation is that aquatic weed compost extracts are better considered stage-sensitive nutrient-biostimulant candidates than direct fertilizer substitutes. Low-dose humic input may be more suitable for seed germination, whereas higher-dose aquatic weed extracts may be more useful for nursery-stage vegetative development. These tendencies require validation with complete replicate-level statistics, improved extract characterization and field-based seedling-to-yield assessment before agronomic recommendation.

ACKNOWLEDGMENTS

This research was funded by the Directorate of Research and Community Service, Ministry of Higher Education, Science, and Technology of the Republic of Indonesia under the Applied Research Prototype Output scheme, with contract number 136/C3/DT.05.00/PL-MULTI TAHUN LANJUTAN/2026, and Number 445/LL9/PPIP.PL-MULTI/2026, 0192/LPPM/II.3.AU/F/202.

AUTHOR CONTRIBUTIONS

Conceptualization, I.R.; Methodology, I.R.; Investigation, J.J.; Data Curation, I.R, H.A., and Y.R.; Writing-Original Draft Preparation, I.R.; Writing Review & Editing, H.A. I.R, S.Z, and Y.R.; Supervision, I.R. All authors have read and agreed to the published version of the manuscript.

CONFLICTS OF INTEREST

The authors declare no conflict of interest.

REFERENCES

- Abbas, S. R., Shahid, I., Javed, M., Malik, K. A., & Mehnaz, S. (2024). Contribution of mineral mobilizing fluorescent pseudomonads in growth promotion of rice (*Oryza sativa* L.) in nutrient deficient soil. *South African Journal of Botany*, 166, 88–96. <https://doi.org/10.1016/j.sajb.2024.01.023>
- Abu-ria, M., Shukry, W., Abo-hamed, S., Albaqami, M., & Almuqadam, L. (2023). Humic Acid Modulates Ionic Homeostasis, Osmolytes Content, and Antioxidant Defense to Improve Salt Tolerance in Rice. *Plants*, 12(1834), 1–20. [source link]
- Adhikari, A., Aneefi, A. G., Sisuvanh, H., Singkham, S., Pius, M. V., Akter, F., Kwon, E.-H., Kang, S.-M., Woo, Y.-J., Yun, B.-W., & Lee, I.-J. (2023). Dynamics of Humic Acid, Silicon, and Biochar under Heavy Metal, Drought, and Salinity with Special Reference to Phytohormones, Antioxidants, and Melatonin Synthesis in Rice. *International Journal of Molecular Sciences*, 24(24). <https://doi.org/10.3390/ijms242417369>

- Asmara, A. P., Sedyadi, E., & Zette, F. I. (2020). Characteristics of liquid product of alkaline treated *Sargassum polycystum* C.A. Agardh. from Lange Beach, Aceh. *Indonesian Journal of Marine Sciences*, 25, 57–65. <https://doi.org/10.14710/ik.ijms.25.2.57-65>
- Bagheri Novair, S., Mirseyed Hosseini, H., Etesami, H., Pirmoradian, N., Asgari Lajayer, B., & Price, G. W. (2024). Straw application improved soil biological properties and the growth of rice plant under low water irrigation. *Environmental Research*, 255. <https://doi.org/10.1016/j.envres.2024.119138>
- Banakar, S. N., Prasannakumar, M. K., Mahesh, H. B., & Parivallal, P. B. (2022). Red-seaweed biostimulants differentially alleviate the impact of fungicidal stress in rice (*Oryza sativa* L.). *Scientific Reports*, 12, 5993. <https://doi.org/10.1038/s41598-022-10010-8>
- Banakar, S. N., Prasannakumar, M. K., Parivallal, P. B., Pramesh, D., Mahesh, H. B., Sarangi, A. N., Puneeth, M. E., & Patil, S. S. (2023). Rice-Magnaporthe transcriptomics reveals host defense activation induced by red seaweed-biostimulant in rice plants. *Frontiers in Genetics*, 14. <https://doi.org/10.3389/fgene.2023.1132561>
- Barus, J., Mustikawati, D. R., Meithasari, D., Ernawati, R., Wardani, N., Tambunan, R. D., & Silalahi, M. (2024). Improvement of growth and nutrient uptake of upland rice grown on degraded acid soil with the application of liquid organic fertilizer. *Journal of Degraded and Mining Lands Management*, 12(1), 6929–6935. <https://doi.org/10.15243/jdmlm.2024.121.6929>
- Begum, S. L. R., Himaya, S. M. M. S., & Afreen, S. M. M. S. (2022). Potential of water hyacinth (*Eichhornia crassipes*) as compost and its effect on soil and plant properties: A review. *Agricultural Reviews*, 43(1), 20–28. <https://doi.org/10.18805/ag.R-184.Submitted>
- Cabrera, E. J. C., Amante, C. P., & Padilla, V. M. (2025). Productivity of Lowland Rice (*Oryza sativa* L.) as Affected by the Integration of Nutrio® Biofertilizer. *Philippine Agricultural Scientist*, 108(2), 171–177. <https://doi.org/10.62550/AQ1501425>
- Canellas, N. A., Olivares, F., & Lopes. (2022). Changes in metabolic profile of rice leaves induced by humic acids. *Plants*, 11, 3261. [source link]
- Chen, H., Yunze, R., & Zhongjun, J. (2025). A Meta-Analysis of 30 Years in China and Micro-District Experiments Shows Organic Fertilizer Quantification Combined with Chemical Fertilizer Reduction Enhances Rice Yield on Saline-Alkali Land. *Rice Science*, 32(2), 259–272. <https://doi.org/10.1016/j.rsci.2025.01.004>
- Choi, S. K., Son, Y. B., Jeong, H. W., & Go, S. (2025). Spatio-temporal projections of the distribution of the canopy-forming algae *Sargassum* in the Western North Pacific under climate change scenarios using the MAXENT model. *Biology*, 14(6), 1–20. [source link]
- Consorti, E., Costarelli, A., Cannavò, S., Cerri, M., Valeri, M. C., Reale, L., Saccomanno, A., Paleni, C., Gregis, V., Kater, M. M., Brilli, F., Paolucci, F., & Ghirardo, A. (2025). Co-Cultivation with *Azolla* Affects the Metabolome of Whole Rice Plant Beyond Canonical Inorganic Nitrogen Fertilization. *Rice*, 18(1). <https://doi.org/10.1186/s12284-025-00788-2>
- Dai, X., Kong, W., Wu, F., Pan, Z., Gao, L., Mo, B., Yu, Y., & Luo, W. (2025). OsNUDX23 regulates early seed germination by modulating ROS balance and starch metabolism in rice. *Frontiers in Plant Science*, 16, 1–16. <https://doi.org/10.3389/fpls.2025.1581800>
- Du, Z., Zhao, P., Fu, Q., Wang, Q., Hu, A., Zhang, W., & Wang, D. (2023). Biostimulants in dissolved organic matters recovered from anaerobic digestion sludge with alkali-hydrothermal treatment: Nontarget identification by ultrahigh-resolution mass spectrometry. *Environment International*, 173. <https://doi.org/10.1016/j.envint.2023.107813>

- Dwi Nurcahyanti, S., Masnilah, R., Budiman, S. A., Tanzil, A. I., & Nurdika, A. A. H. (2024). Microbial consortium formulation in liquid organic fertilizer for managing bacterial leaf blight (*Xanthomonas oryzae* pv. *Oryzae*), rice blast (*Pyricularia oryzae*), and enhancing rice productivity. *Biodiversitas*, 25(5), 2209–2220. <https://doi.org/10.13057/biodiv/d250539>
- Fitriyah, F., Aziz, M. A., Wahyuni, S., Fadila, H., & Meida, I. (2022). Biostimulant activity of *Sargassum* sp. extracts on early growth of *Zea mays* L. and the phytohormones content analysis. *JTBB*, 7(2), 1–13. <https://doi.org/10.22146/jtbb.69178>
- Guidoni, L. L. C., Martins, G. A., Guevara, M. F., Brandalise, J. N., Lucia, T., Gerber, M. D., Corrêa, L. B., & Corrêa, É. K. (2021). Full-Scale Composting of Different Mixtures with Meal from Dead Pigs: Process Monitoring, Compost Quality and Toxicity. *Waste and Biomass Valorization*, 12(11), 5923–5935. <https://doi.org/10.1007/s12649-021-01422-0>
- Hamouda, M. M., Saad, K. M., & Dina, A. (2022). Potential of seaweed extract on growth, physiological, cytological and biochemical parameters of wheat (*Triticum aestivum* L.) seedlings. *Journal of Soil Science and Plant Nutrition*, 22, 1818–1831. <https://doi.org/10.1007/s42729-022-00774-3>
- Haque, M. M., Datta, J., Ahmed, T., Ehsanullah, M., Karim, M. N., Akter, M. S., Iqbal, M. A., Baazeem, A., Hadifa, A., Ahmed, S., & Sabagh, A. E. L. (2021). Organic amendments boost soil fertility and rice productivity and reduce methane emissions from paddy fields under sub-tropical conditions. *Sustainability (Switzerland)*, 13(6). <https://doi.org/10.3390/su13063103>
- Ibraheem, F., Abu-Ria, M. E., Abo-Hamed, S. A., Anis, G. B., & Shukry, W. M. (2023). Humic Acid Improves Germination, Vegetative Growth, and Yield in Rice (*Oryza sativa* L.) Under Salinity Stress. *Catrina -The International Journal of Environmental Sciences*, 27(1), 75–91. <https://doi.org/10.21608/cat.2023.201982.1164>
- Indriani, T., Bertham, Y. H., Hasanudi, Prasetyo, & Simanihuruk, B. W. (2022). Effect of humic acid on nutrient availability and yield of upland rice inoculated with biofertilizers in the coastal area. *TERRA*, 5(1), 15–20. [source link]
- Iriany, A., Prihartini, I., Zakia, A., Septia, E. D., Farahdina, F. A. R., & Hasanah, F. (2023). Effects of Different Bioactivator Rates on Growth, Yield and Soil Properties of Two Rice Varieties in Indonesia. *International Journal of Design and Nature and Ecodynamics*, 18(5), 1151–1157. <https://doi.org/10.18280/IJDNE.180516>
- Ishaq, S., Belduz, A. O., Ceylan, E., Senocak, A. N., Munawar, W., Alkowlani, A. T. H., Terzi, R., Bektas, K. I., & Canakci, S. (2025). Plant growth-promoting bacteria from Uzungöl forest stimulate rice growth via seed biopriming and root inoculation: Isolation and functional characterization of potent PGPR strains from rhizosphere soils of different trees. *Frontiers in Plant Science*, 16. <https://doi.org/10.3389/fpls.2025.1622951>
- Izquierdo, J., Arriagada, O., García-Pintos, G., Ortiz, R., García-Pintos, M., & García-Pintos, M. (2025). Humic field biostimulation as a sustainable agricultural practice to increase yield of main grains: Evidence from on-farm trials. *Frontiers in Plant Science*, 16. <https://doi.org/10.3389/fpls.2025.1709876>
- Ji, R., Liu, C., Xu, Q., Zhang, Y., Chen, M., Zhang, L., & Hu, F. (2024). Effect of Artificial Humic Acids Derived from Municipal Sludge on Plant Growth, Soil Fertility, and Dissolved Organic Matter. *Agriculture (Switzerland)*, 14(11). <https://doi.org/10.3390/agriculture14111946>
- Kalita, N., Boruah, P., Maibangsa, S., Bhuyan, S., Gogoi, B., & Saud, R. K. (2025). Response of Transplanted Rice to Seedling Root-dip in Phosphorus and Biofertilizer Slurry in Acid Soils of North East India- A Participatory Assessment in Farmers' Field. *Indian Journal of Agricultural Research*, 59(5), 719–724. <https://doi.org/10.18805/IJARE.A-6127>

- Kanga, C. N., Umezawa, R., Saha, S. R., Takahashi, H., Yamasaki, M., & Itoh, K. (2026). Volatile Compounds from Waste Shiitake Fungi Beds Enhance Rice Growth, Yield, and Performance Under High-Temperature Field Conditions. *Agronomy*, 16(9). <https://doi.org/10.3390/agronomy16090892>
- Kaur, P., Saini, K. S., Sharma, S., Kaur, J., Bhatt, R., Alamri, S., Alfagham, A. T., & Hussain, S. (2023). Increasing the Efficiency of the Rice–Wheat Cropping System through Integrated Nutrient Management. *Sustainability (Switzerland)*, 15(17). <https://doi.org/10.3390/su151712694>
- Kumar, S. R., David, E. M., Pavithra, G. J., Kumar, G. S., Lesharadevi, K., Akshaya, S., Basavaraddi, C., Navyashree, G., Arpitha, P. S., Sreedevi, P., Zainuddin, K., Firdous, S., Babu, B. R., Prashanth, M. U., Ravikumar, G., Basavaraj, P., Chavana, S. K., Kumar, V. M. L. D., Parthasarathi, T., & Subbian, E. (2024). Methane-derived microbial biostimulant reduces greenhouse gas emissions and improves rice yield. *Frontiers in Plant Science*, 15. <https://doi.org/10.3389/fpls.2024.1432460>
- Kurniawan, L., Maryudi, M., & Astuti, E. (2024). Utilization of tofu liquid waste as liquid organic fertilizer using the fermentation method with activator effective microorganisms 4 (EM-4): A review. *Equilibrium Journal of Chemical Engineering*, 8(1), 100–112. [source link]
- Li, Z., Henawy, A. R., Halema, A. A., Fan, Q., Duanmu, D., & Huang, R. (2022). A Wild Rice Rhizobacterium *Burkholderia cepacia* BRDJ Enhances Nitrogen Use Efficiency in Rice. *International Journal of Molecular Sciences*, 23(18). <https://doi.org/10.3390/ijms231810769>
- Linda, T. M., Aliska, J., Feronika, N., Melisa, I., & Juliantari, E. (2024). Production of Exopolysaccharides and Indole Acetic Acid (IAA) by Rhizobacteria and Their Potential against Drought Stress in Upland Rice. *Journal of Microbiology and Biotechnology*, 34(6), 1239–1248. <https://doi.org/10.4014/jmb.2401.01035>
- Liu, L., Yang, X., Robakowski, P., Ye, Z., Wang, F., & Zhou, S. (2025). Mechanistic modeling reveals adaptive photosynthetic strategies of *Pontederia crassipes*: Implications for aquatic plant physiology and invasion dynamics. *Biology*, 14(6), 1–15. [source link]
- Liu, T., Liu, Z., Zhao, Z., Xu, K., Chen, H., Feng, Y., Wang, W., Zhang, N., Liu, D., He, X., & Wu, J. (2023). Low-Temperature Fermented Straw Compost Regulates Rice Growth and Yield by Affecting Soil Physicochemical Properties and the Expression of Important Signaling Pathway Genes. *Agronomy*, 13(12). <https://doi.org/10.3390/agronomy13123066>
- Luo, L., Zhang, Y., & Xu, G. (2020). How does nitrogen shape plant architecture? *Journal of Experimental Botany*, 71(15), 4415–4427. <https://doi.org/10.1093/jxb/eraa187>
- Marti-Jerez, K., Català-Forner, M., Tomàs, N., Murillo, G., Ortiz, C., Sánchez-Torres, M. J., Vitali, A., & Lopes, M. S. (2023). Agronomic performance and remote sensing assessment of organic and mineral fertilization in rice fields. *Frontiers in Plant Science*, 14. <https://doi.org/10.3389/fpls.2023.1230012>
- Minhas, A., Saeed, M. S., Ehsan, A., Murtaza, G., Akram, M., Mehran, M., Aurangzeb, M., Elahi, N. N., Iqbal, R. K., & Syed, A. (2024). RESPONSE OF INTEGRATED USE OF HUMIC ACID AND CHEMICAL FERTILIZER ON GROWTH AND YIELD OF RICE CROP (*ORYZA SATIVA* L.) IN CALCAREOUS SOIL. *Pakistan Journal of Botany*, 56(3), 879–887. [https://doi.org/10.30848/PJB2024-3\(41\)](https://doi.org/10.30848/PJB2024-3(41))
- Monajjem, S., Soltani, E., Zainal, E., Esfahani, M., Ghaderi-Far, F., Chaleshtori, M. H., & Rezaei, A. (2023). Seed priming improves enzymatic and biochemical performances of rice during seed germination under low and high temperatures. *Rice Science*, 30(4), 335–347. <https://doi.org/10.1016/j.rsci.2023.03.012>

- Naveen, P. V., Maitra, S., Chhetri, P., Sairam, M., Dash, B., & Ray, S. (2026). Effect of Bio-Stimulant and Nitrogen Levels on Growth and Productivity of Kharif Rice. *Plant Science Today*, 13(1), 1–6. <https://doi.org/10.14719/pst.10681>
- Prasedya, E. S., Kurniawan, N. S. H., Fitriani, F., Saraswati, P. B. A., Qoriasmadillah, W., Ilhami, B. T. K., Hernawan, A., & Widyastuti, S. (2023). Sustainable Use of Organic Seaweed Fertilizer Improves the Metagenomic Function of Microbial Communities in the Soil of Rice Plants. *Sustainability (Switzerland)*, 15(23). <https://doi.org/10.3390/su152316328>
- Qi, Z., Xu, C., Tang, R., Zhang, Q., Sun, W., Guan, C., & Wang, Y. (2025). Response of photosynthesis and chlorophyll fluorescence to nitrogen changes in rice with different nitrogen use efficiencies. *Plants*, 14, 1–18. [source link]
- Riddech, N., Ngo, M. N., Boonlue, S., Dongsansuk, A., Santanoo, S., & Theerakulpisut, P. (2025). Physical and Chemical Properties Evaluation of Liquid Organic Fertilizers (LOFs) and Their Effects on Promoting Rice Growth. *Sustainability (Switzerland)*, 17(7). <https://doi.org/10.3390/su17073087>
- Ros, M., Hurtado-Navarro, M., Giménez, A., Egea-Gilabert, C., Lozano-Pastor, P., & Pascual, A. (2020). Spraying agro-industrial compost tea on baby spinach crops: Evaluation of yield, plant quality and soil health in field experiments. *Agronomy*, 10(3), 1–15. [source link]
- Rosawanti, P. (2019). Kandungan unsur hara pada pupuk organik tumbuhan air lokal. *Jurnal Daun*, 6(2), 140–148. [source link]
- Rozen, N., Kasim, M., Kusumawati, A., Sholihat, I., Ikram, H., & Gusril, A. (2024). Application of Liquid Organic Fertilizer to Improve Yields in System of Rice Intensification (SRI). *Pakistan Journal of Biological Sciences*, 27(7), 365–372. <https://doi.org/10.3923/pjbs.2024.365.372>
- Selvam, S., & Kumar, K. S. (2022). Application of banana spathe extracts and compost for improving growth in rice plants. *Journal of Applied Biology and Biotechnology*, 10(1), 112–119. <https://doi.org/10.7324/JABB.2021.100114>
- Senthilkumar, S., Kuppusamy, S., Palai, S., Rajanbabu, V., & A., V. (2024). Exploring Sargassum extract as a bio-stimulant: Advancements and benefits in soil-plant systems—A review. *Communications in Soil Science and Plant Analysis*, 55, 3638–3648. <https://doi.org/10.1080/00103624.2024.2397013>
- Shultana, R., Zuan, A. T. K., Rana, M., Naher, U. A., Paul, P. L. C., Akter, M., Shupta, S. A., & Roy, T. K. (2025). Exploring indigenous *Bacillus* spp. As a biostimulant to enhance the growth and yield of rice under glasshouse conditions. *Asian Journal of Agriculture*, 9(1), 131–139. <https://doi.org/10.13057/asianjagrig/g090114>
- Thaimei, T., Bokado, K., Meetei, T. T., Khwairakpam, R., & Bera, B. (2025). Effect of different level of poultry manure and seaweed extract on growth, yield and quality of rice (*Oryza sativa* L.). *Journal of Applied and Natural Science*, 17(4), 1877–1885. <https://doi.org/10.31018/jans.v17i4.7021>
- Umamaheswari, J., & Shanthakumar, S. (2021). Paddy-soaked rice mill wastewater treatment by phycoremediation and feasibility study on use of algal biomass as biofertilizer. *Journal of Chemical Technology and Biotechnology*, 96(2), 394–403. <https://doi.org/10.1002/jctb.6551>
- Utomo, W., & Guntoro, D. (2023). Potensi ekstrak daun eceng gondok (*Eichhornia crassipes* (Mart.) Solms-Laub.) sebagai bioherbisida untuk mengendalikan gulma pada padi sawah. *Buletin Agrohorti*, 11(1), 136–142. [source link]
- Vaishnavi, D., Ravichandran, V., Sritharan, N., Anandakumar, S., Prabu Kumar, G., & Latha, M. R. (2024). Biopriming of seeds with microbial biostimulant (*Bacillus megaterium*) on improvement

- of seedling growth, biochemical and root traits of rice (*Oryza sativa* L.). *Plant Science Today*, 11. <https://doi.org/10.14719/pst.5541>
- Wang, Q., Li, S., Li, J., & Dong, H. (2024). The utilization and roles of nitrogen in plants. *Forests*, 15(7), 1–20. [source link]
- Xie, H., Wu, K., Iqbal, A., Ali, I., He, L., Ullah, S., Wei, S., Zhao, Q., Wu, X., Huang, Q., & Jiang, L. (2021). Synthetic nitrogen coupled with seaweed extract and microbial inoculants improves rice (*Oryza sativa* L.) production under a dual cropping system. *Italian Journal of Agronomy*, 16(2). <https://doi.org/10.4081/ija.2021.1800>
- Zega, I. C., Mendrofa, Y. T., Mendrofa, E., & Ndraha, A. B. (2025). Pemanfaatan eceng gondok (*Eichhornia crassipes*) dalam meningkatkan kesuburan tanah. *Jurnal Kajian Ilmu Pertanian dan Perkebunan*, 2(1), 161–167. [source link]
- Zhang, S., Tang, L., Zhan, X., Wang, D., Zhang, A., Wu, H., Huang, C., Chen, H., & Wang, J. (2025). Elucidating the mechanism of soybean-derived protein hydrolysate in stabilizing rice yield and enhancing agronomic efficiency. *Frontiers in Plant Science*, 16. <https://doi.org/10.3389/fpls.2025.1651406>